

Systems Biology
Linking all the components of a cell in a quantitative and temporal manner

Origins of Cellular Complexity

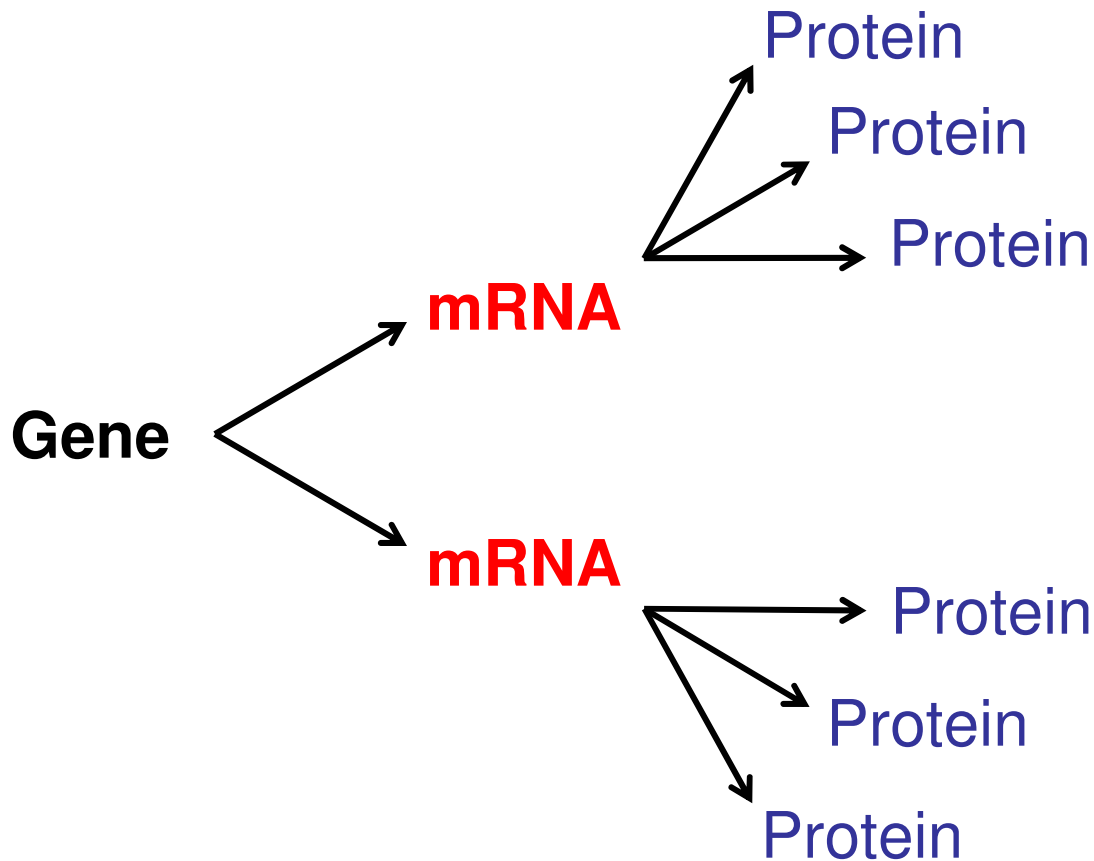
Humans

C. elegans

<ul style="list-style-type: none">• ~3,000 megabases• ~21,000-25,000 Genes• ~10,000,000,000,000 cells	<ul style="list-style-type: none">• ~100 megabases• ~20,000 Genes• ~1,000 cells
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- Alternative Splicing
- Improved and modular interaction between components
- Improved spatial relationship between components in the cell
- Improved interaction between intracellular signaling pathways
- Multiple protein forms and functions
- etc...





- Covalent Modification or Modifications
 - Phosphorylation
 - Glycosylation
 - Lipidation
 - Nitrosylation
 - Acetylation, etc...
- Cellular localization
 - Nucleous
 - Mitochondria
 - Endoplasmic Reticulum, etc...
- Presence or Absence of Ligands
 - Heme
 - Metal
 - ATP, etc...
- Proteolytic Cleavage
 - N-terminal and C-terminal truncations
- Oligomeric State
 - ~78% of yeast proteins act in part of a greater complex of proteins
- Protein Conformation

What needs to be measured?

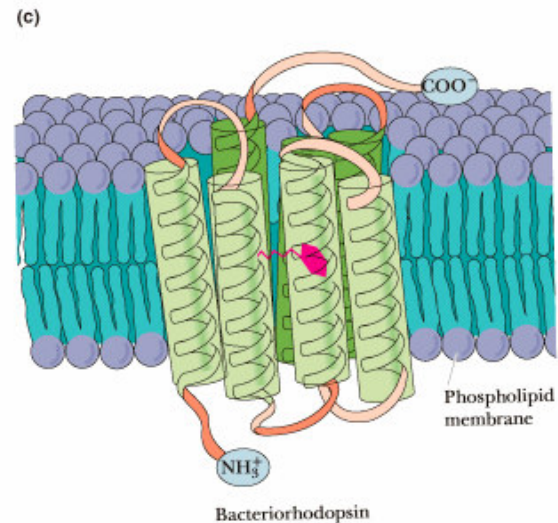
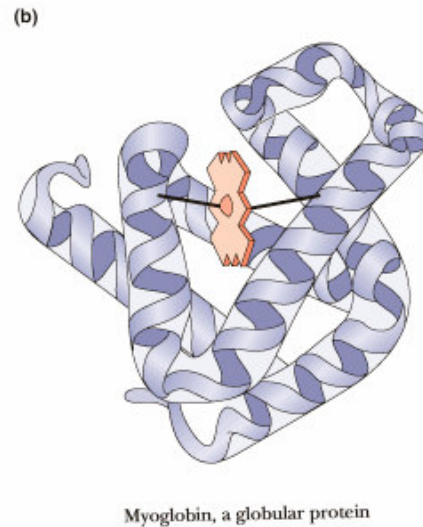
- Need methods that can detect and quantify all proteins (and modifications)
- Determine protein-protein interactions
- Determine protein localization
- Assess protein function
- Evaluate conformational changes

At X points in time and under Y different conditions

Must have methods that can measure more than one protein/modification/interaction/etc... at a time!!!!

Dealing with protein is difficult!

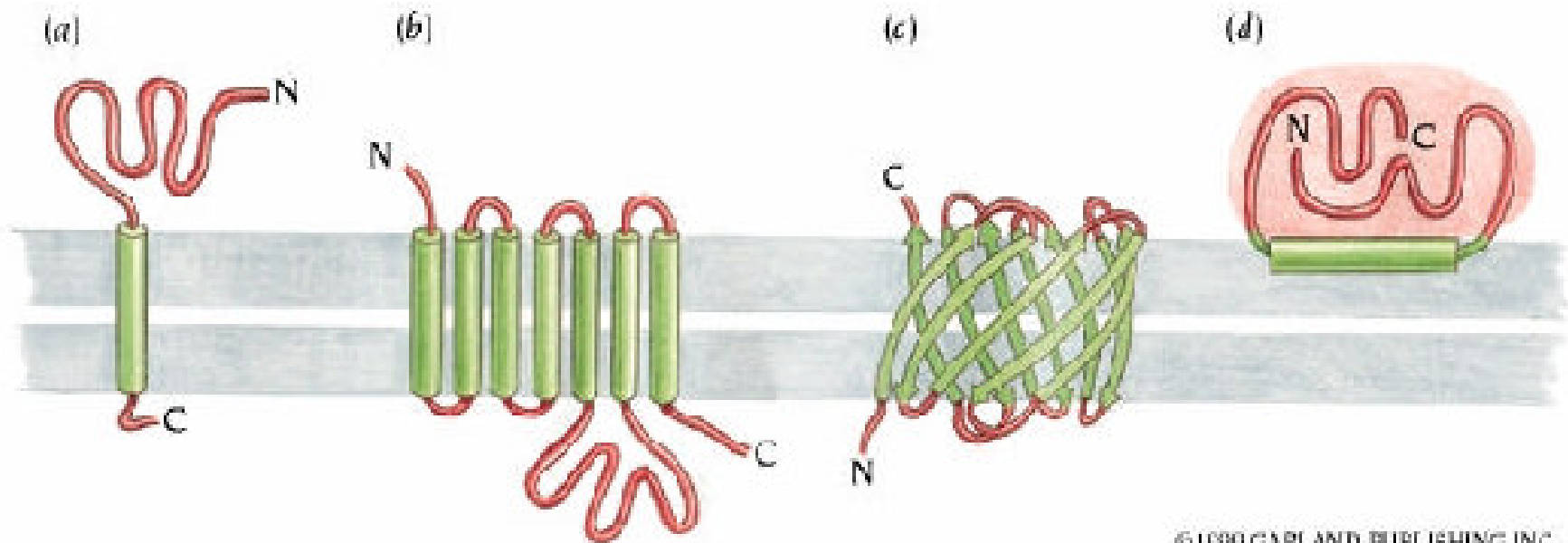
Proteins span an enormous range of physiochemical properties



- 25-30% of all Eukaryote Proteins
- 50% of all drug targets
- Insoluble in aqueous solutions

~90% of all analyses are focused on soluble proteins!!

Arrangement of Proteins in a Lipid Bilayer



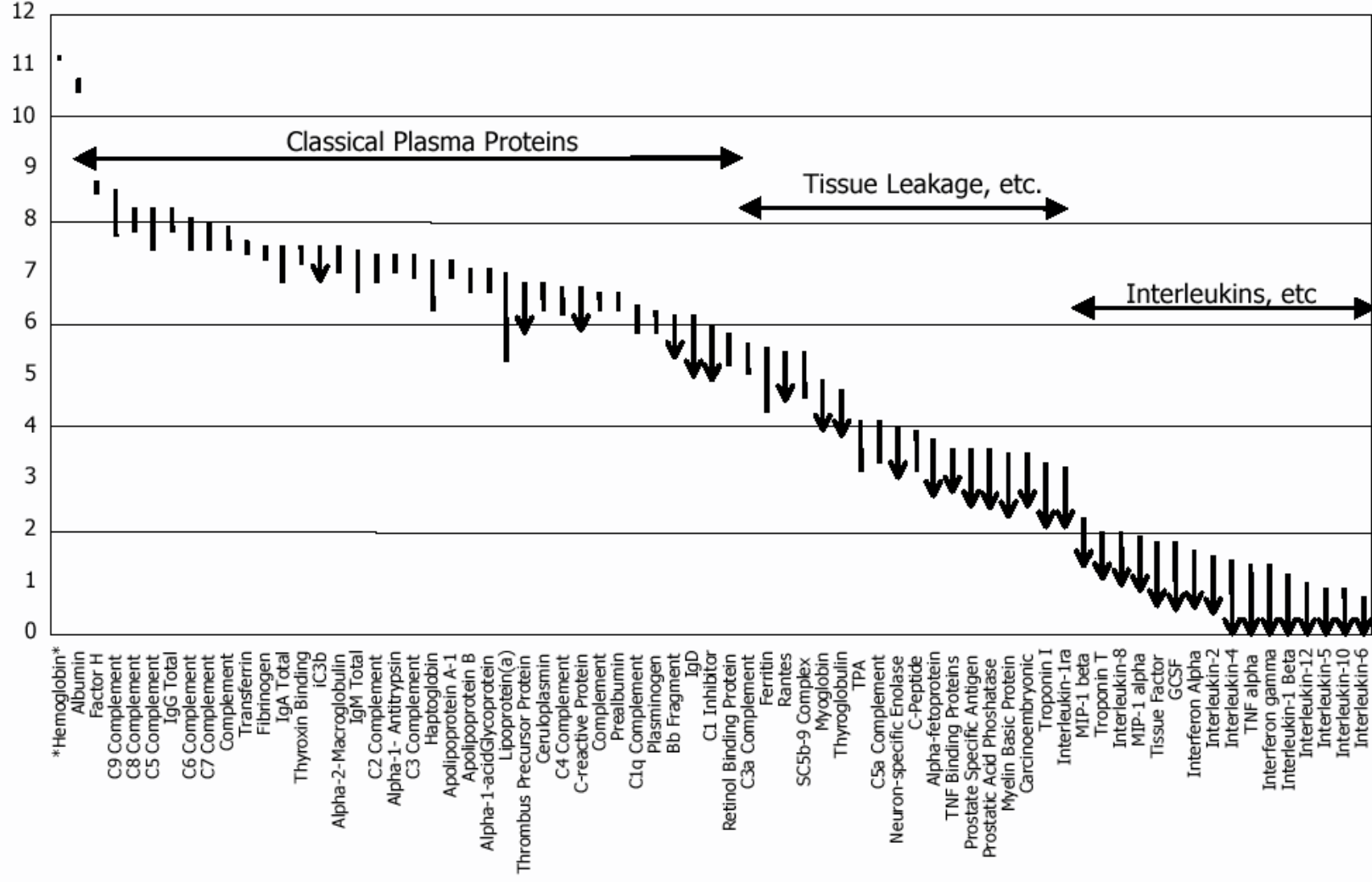
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The Dynamic Range and Sensitivity is a Problem

Table 1. Detection limits for proteins assuming 100% recovery

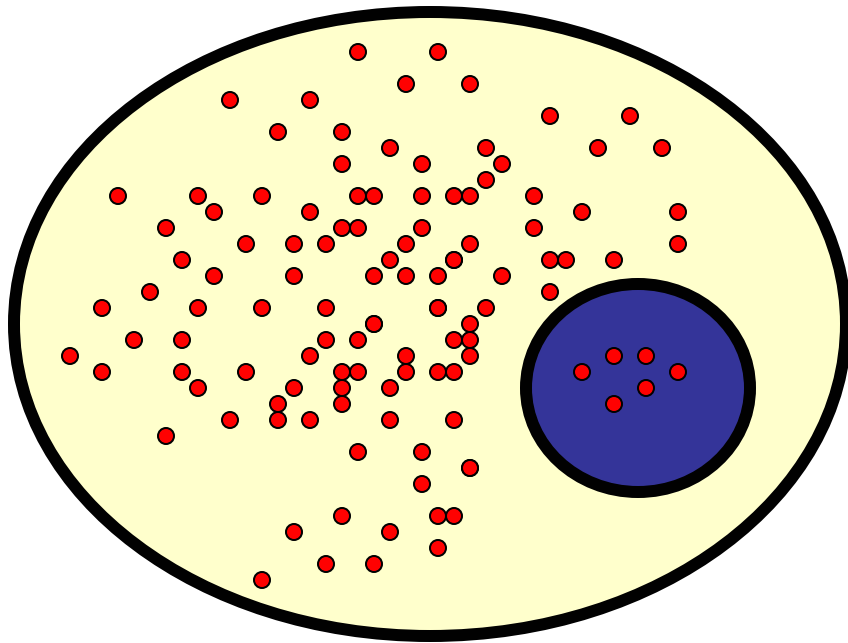
Number of cells	Protein copies/cell	Total number of molecules	Moles of protein
1.OE+09	1000000	1.OE+15	1600 pmole
1.OE+09	100000	1.OE+14	160 pmole
1.OE+09	10000	1.OE+13	16 pmole
1.OE+09	1000	1.OE+12	1.6 pmole
1.OE+09	100	1.OE+11	160 fmole
1.OE+09	10	1.OE+10	16 fmole
1.OOE+08	1000000	1.OE+14	160 pmole
1.OOE+08	100000	1.OE+13	16 pmole
1.OOE+08	10000	1.OE+12	1.6 pmole
1.OOE+08	1000	1.OE+11	160 fmole
1.OOE+08	100	1.OE+10	16 fmole
1.OOE+08	10	1.OE+09	1.6 fmole
1.OOE+07	1000000	1.OE+13	16 pmole
1.OOE+07	100000	1.OE+12	1.6 pmole
1.OOE+07	10000	1.OE+11	160 fmole
1.OOE+07	1000	1.OE+10	16 fmole
1.OOE+07	100	1.OE+09	1.6 fmole
1.OOE+07	10	1.OE+08	0.2 fmole
1.OOE+06	1000000	1.OE+12	1.6 pmole
1.OOE+06	100000	1.OE+11	160 fmole
1.OOE+06	10000	1.OE+10	16 fmole
1.OOE+06	1000	1.OE+09	1.6 fmole
1.OOE+06	100	1.OE+08	0.2 fmole

Normal Range Abundances Log₁₀ Concentration in pg/mL

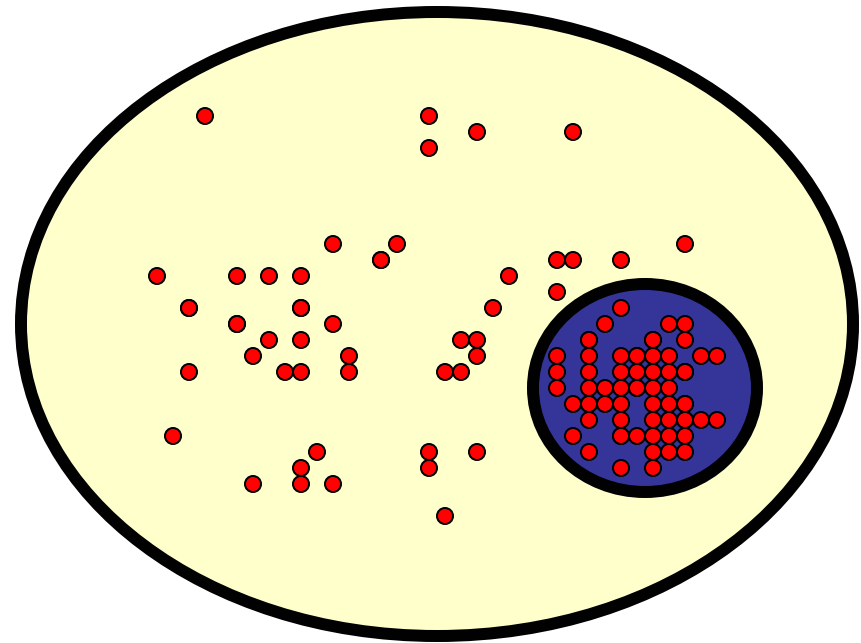


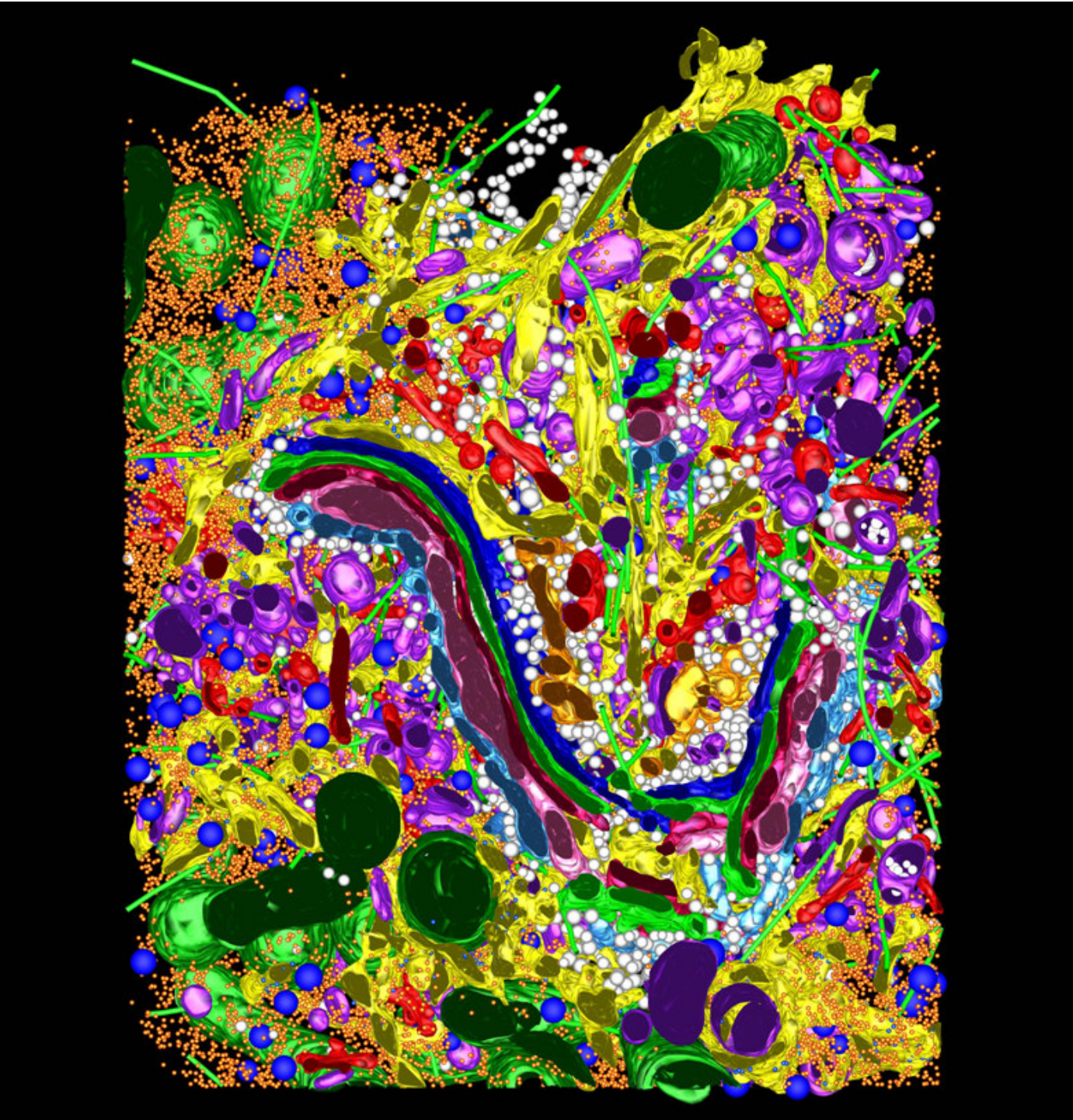
We have a protein in multiple compartments and the localized level changes but not the overall cellular level. How do we measure this?

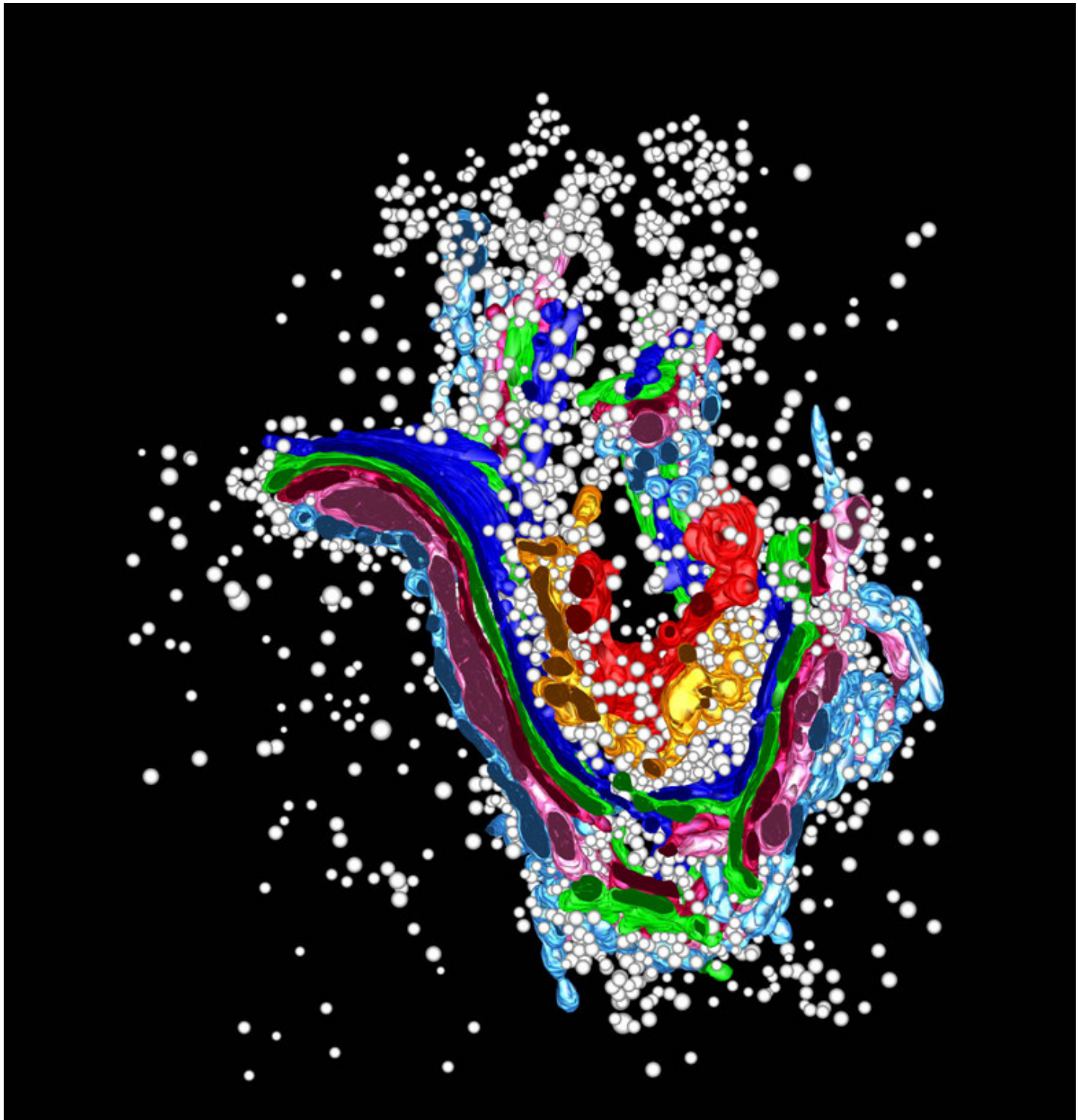
1.



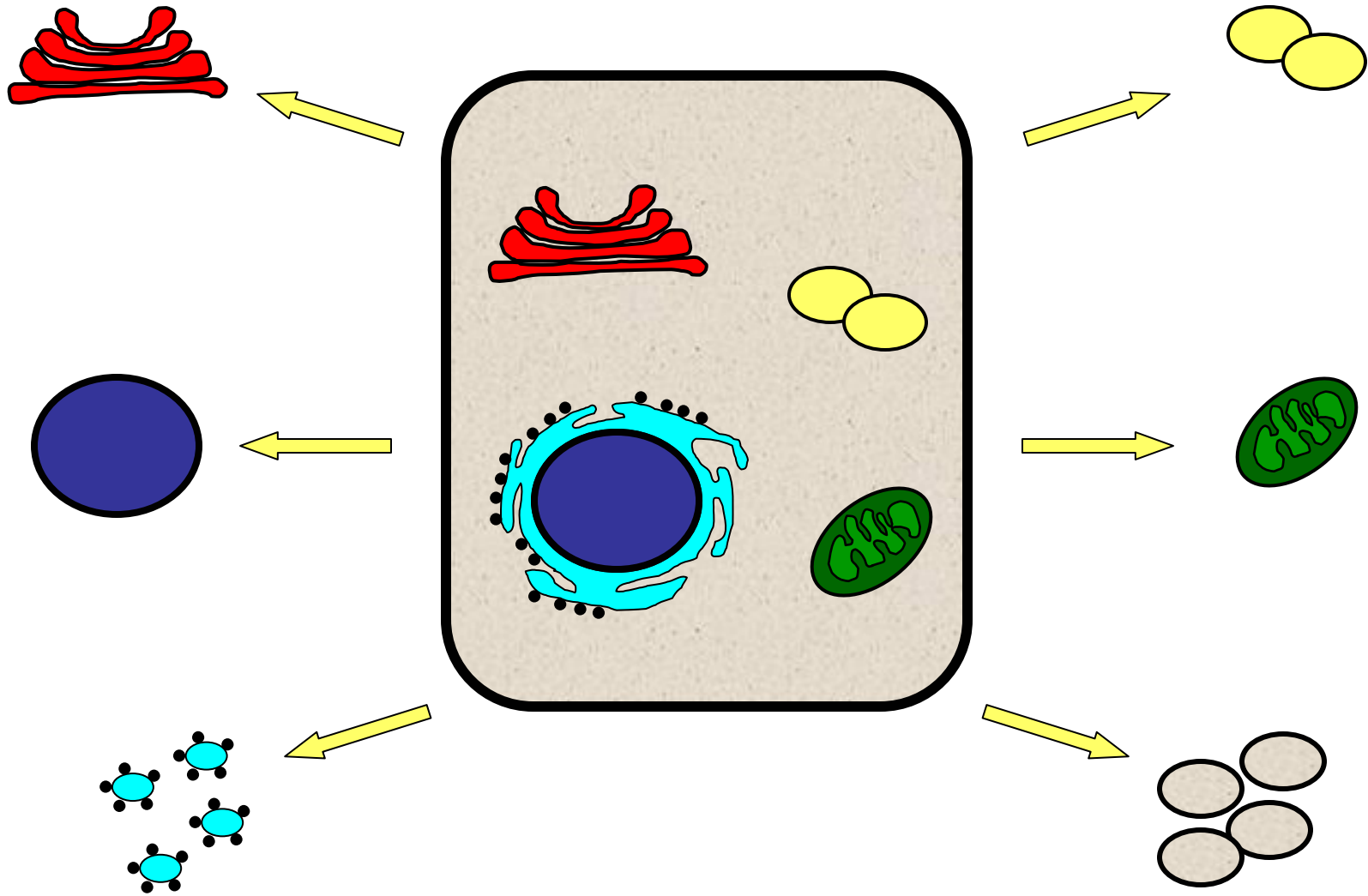
2.





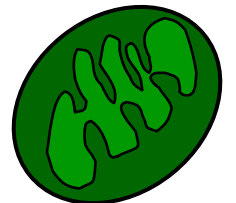


Cell Fractionation

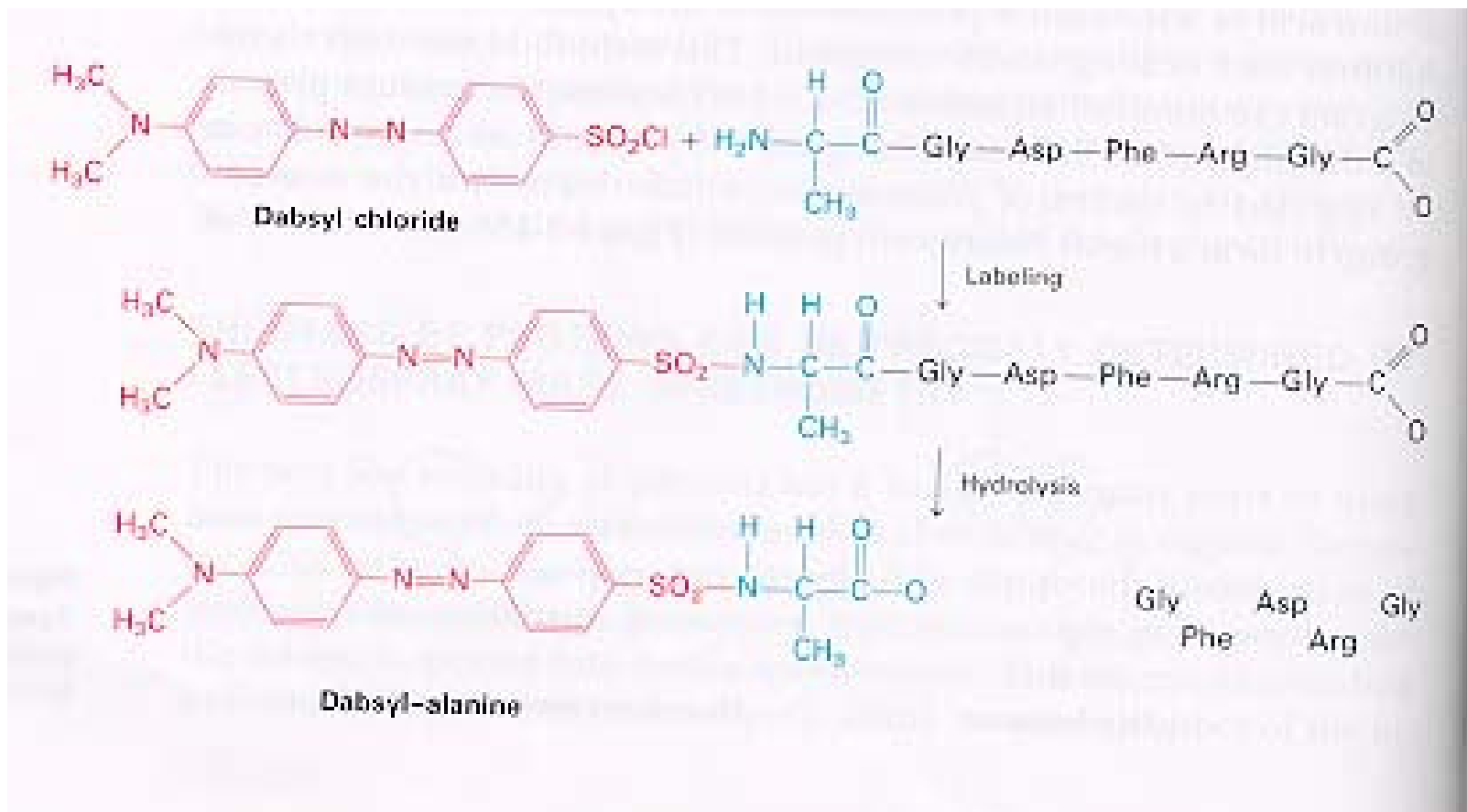


How do we distinguish between a high abundance contaminant or a low abundance specific protein?

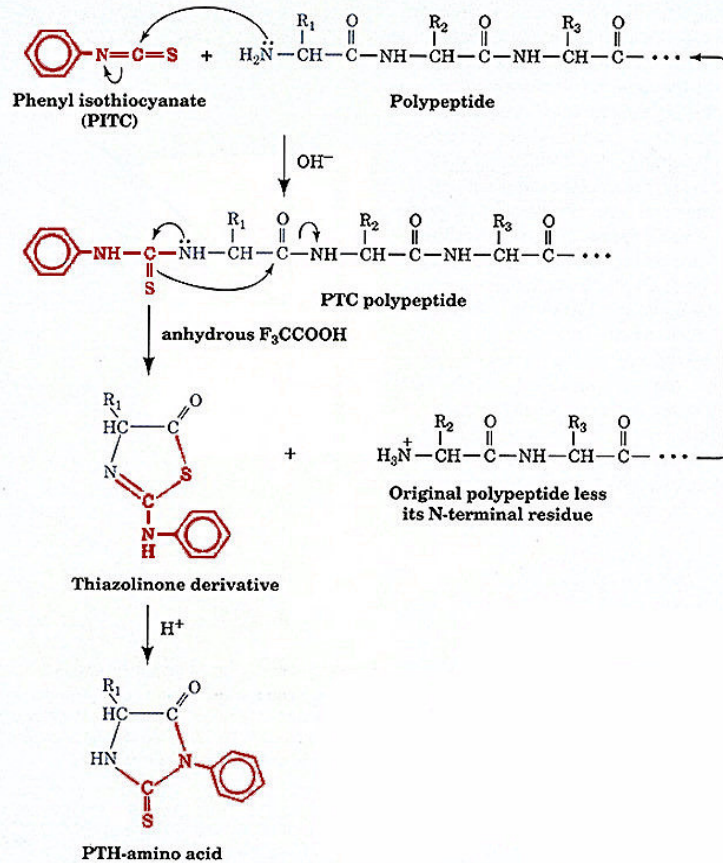
- Consider two proteins present in a cell.
 - Protein A is localized to the cytosol and is present at 1,000,000 copies/cell
 - Protein B is localized to the mitochondria and is present 100 copies/cell
- If the mitochondria is then enriched 100x
 - Protein A will be depleted 100x => 10,000
 - Protein B will be enriched 100x => 10,000



N-terminal amino acid can be identified by reaction with dabsyl chloride or dansyl chloride followed by acid hydrolysis



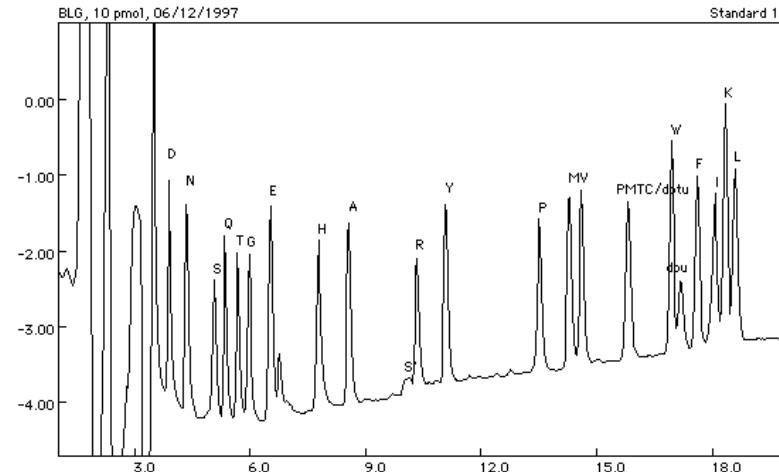
N-Terminal Protein Sequencing



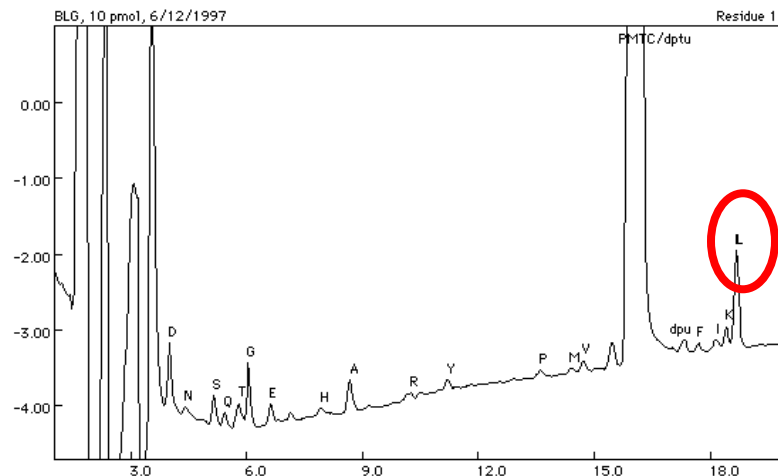
30-50 min per cycle
Each cycle is ~98% effective.
Capable of sequencing ~50 AA.

Edman P. A method for determination of the amino acid sequence in peptides, *Arch. Biochem.* **22**: 475-476 (1949)

Standard



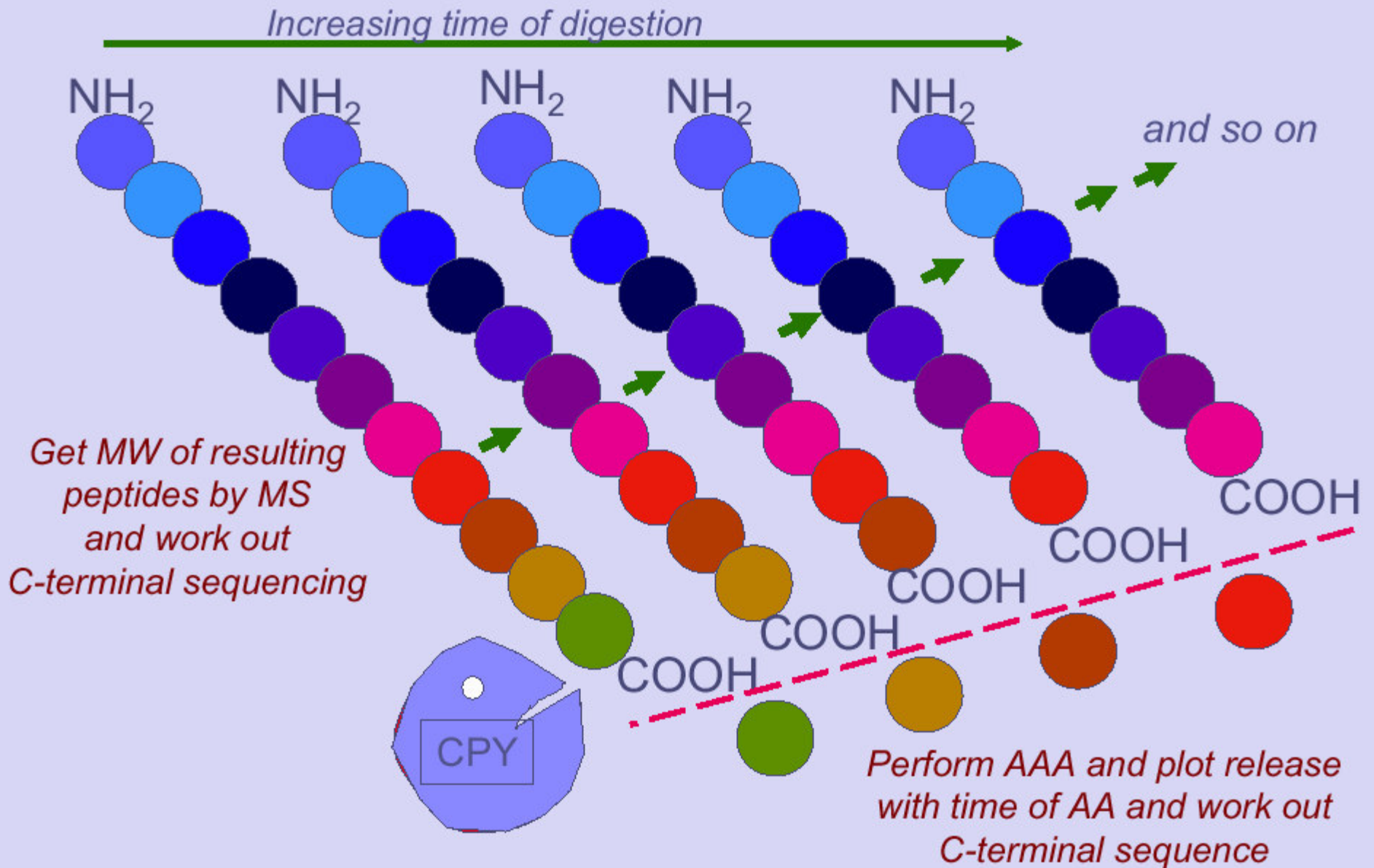
Residue 1



C-terminal Methods are Almost Non Existent

- Most methods involve aminopeptidases (carboxypeptidases) to selectively cleave one amino acid at a time from the C-terminus.
 - This amino acid is then determined by chromatographic methods or
 - remaining protein sequence is measured by mass spectrometry.

C-terminal Sequencing by MS



Determining the Primary Structure of a Protein

- Protein sequencing has become**
- largely obsolete because of large**
- scale DNA sequencing and**
- protein coding gene predictions.**
- Most strategies now focus on**
- detection instead of sequencing.**

Post-Translational Modifications

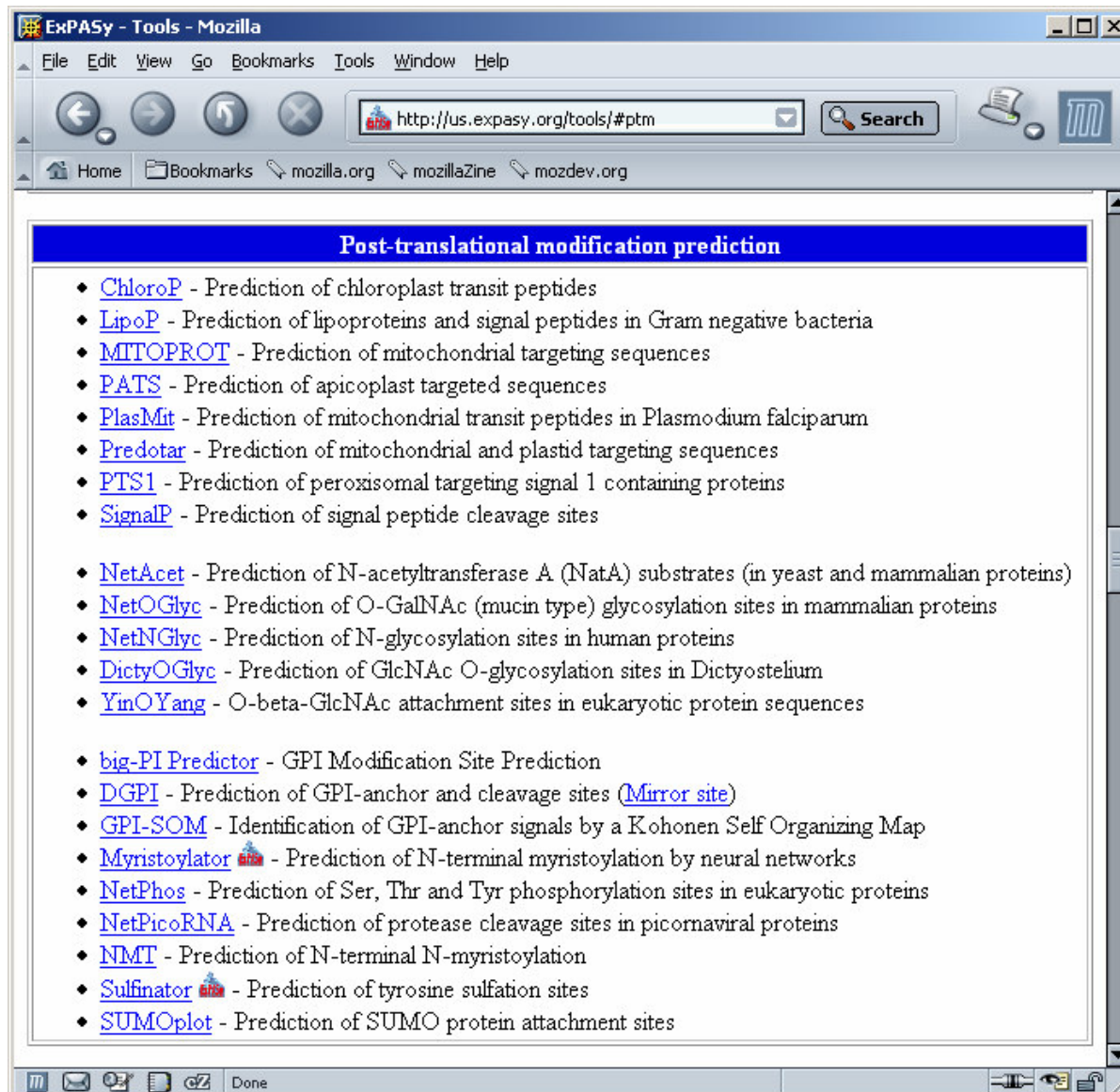
<http://www.abrf.org/index.cfm/dm.home>

351 Modifications in Delta Mass. Not all of these occur *in vivo*

The screenshot shows a Mozilla browser window titled "ABRF Delta Mass Reference - Mozilla". The address bar contains the URL <http://www.abrf.org/index.cfm/dm.home>. The browser's menu bar includes File, Edit, View, Go, Bookmarks, Tools, Window, and Help. The page content features a search bar with a "GO" button and a "Search" button. A navigation menu on the left lists: About ABRF, Forms and Documents, News and Announcements, Communications, Research Groups, Open Research Studies, Committees, Reference, and Sponsorship. The main heading is "Delta Mass" in large blue font, followed by the subtitle "A Database of Protein Post Translational Modifications" in green. Below this is a link to "Contributors" in blue. A light blue box on the right contains two portrait photos of men and a dedication: "This page is dedicated to the memories of [Finn Wold](#) and [Ken Mitchelhill](#)". A blue notice states: "The ABRF accepts no responsibility for the accuracy of these data which are freely donated by individuals". At the bottom, there is a search section titled "Search the Delta Mass Database" with a "View All" link. It includes instructions: "Enter a +/- integer to search on, and select a margin of error (range of search)." and a search form with fields for "Avg. Mass Change", "Error Margin" (set to "Exact"), and a "Search!" button. The status bar at the bottom indicates "Transferring data from www.abrf.org...".

Post-Translational Modification Site Prediction

<http://us.expasy.org/tools/#ptm>



Mostly stores motifs.

For Example:
Protein kinase C (PKC)
recognizes [ST]-x-[RK]

Thus any protein with:

S-X-R

T-X-R

S-X-K

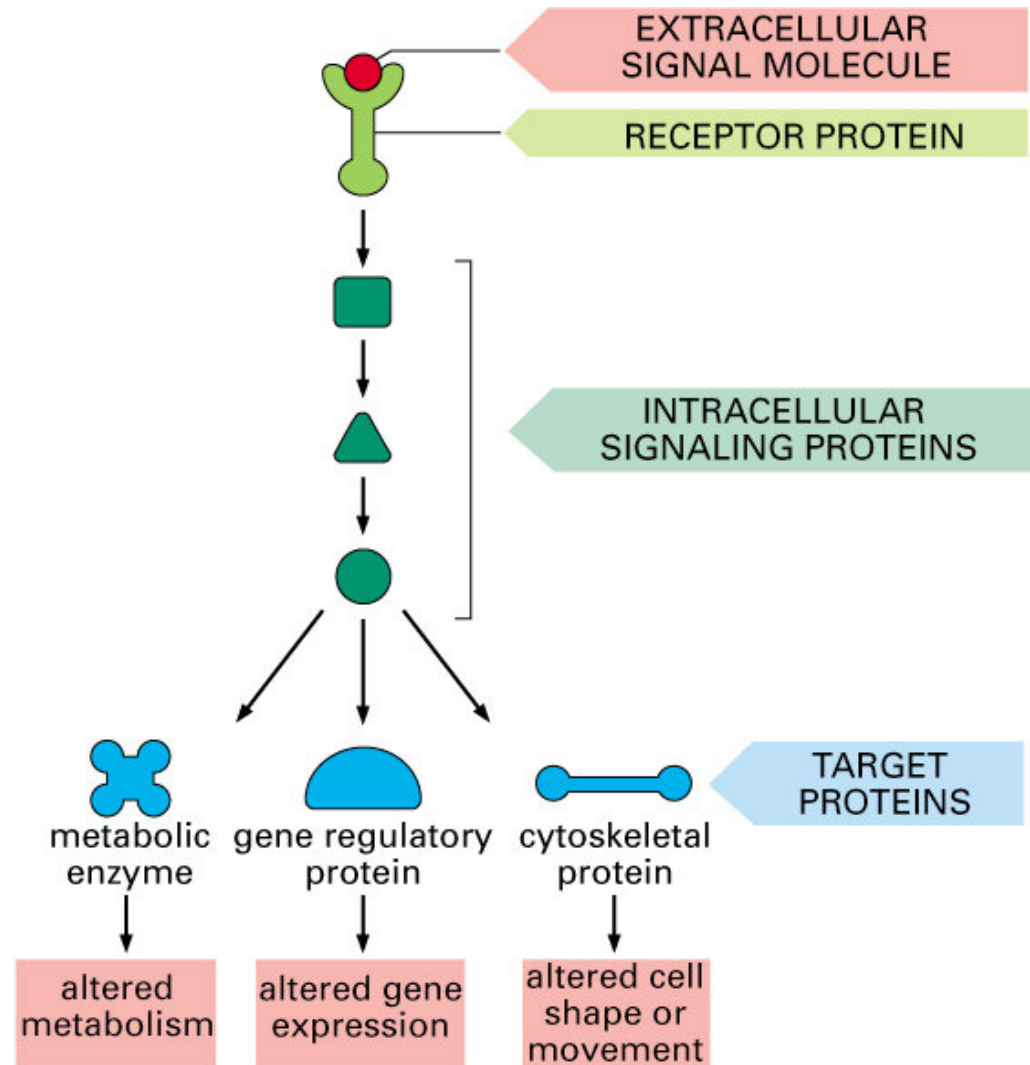
T-X-K

Would come back with a
predicted phosphorylation
site.

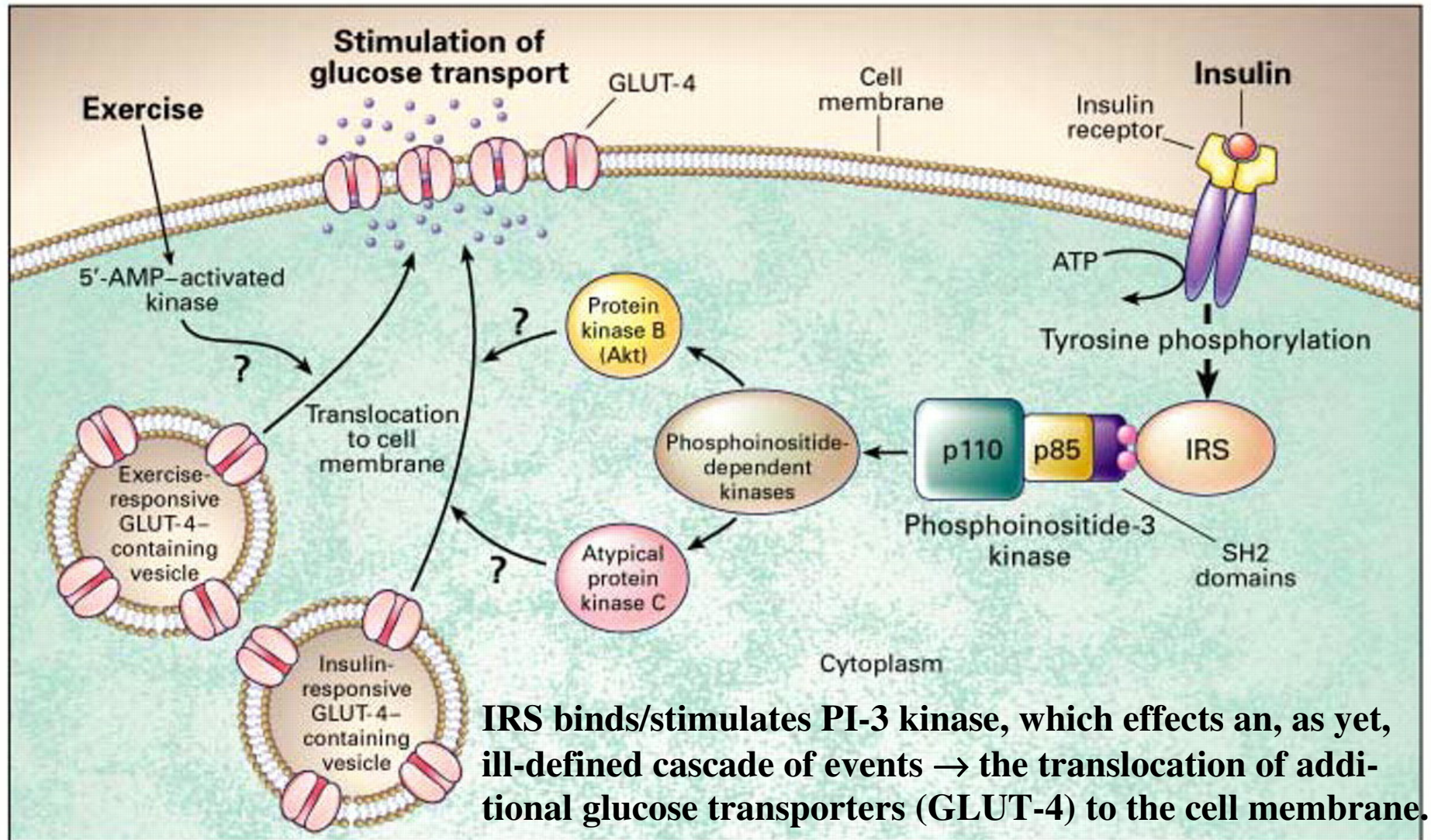
INTRACELLULAR SIGNAL TRANSDUCTION: A Journey from the Plasma Membrane to the Nucleus

How do we determine the components?

How do we determine the mechanism of action of a signaling pathway?



Example: Insulin Stimulation of Glucose Transport



New Engl J Med 341:248, 1999

GLUT-4 Translocation to the Plasma Membrane

How do we determine the substrates for a kinase, ubiquitin ligase, methyl transferase?

What if the stoichiometry of the modification is low?

What if the phosphorylation / dephosphorylation event is very rapid?

How would we know it even happened?